

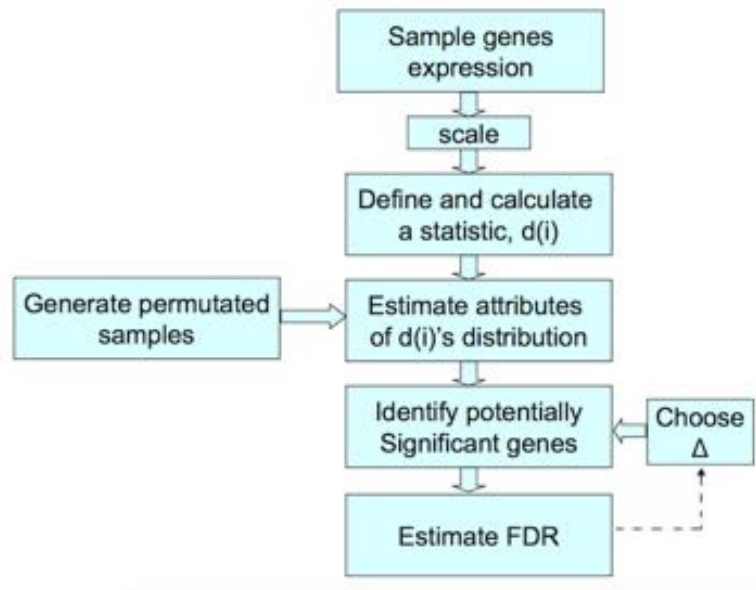
Significance Analysis for Microarrays

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Significance analysis of microarrays (SAM)

- V. G. Tusher et.al. "Significance Analysis of Microarrays Applied to the Ionizing Radiation Response" PNAS 2001
<http://www.pnas.org/content/98/9/5116.long>
- A clever adaptation of the t-ratio to borrow information across genes
- SAM seeks to control the proportion of false rejections among the set of rejected hypotheses (FDR).
- Permutation method is used to calculate the null distribution of the modified t-statistics.

SAM procedure



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SAM t-test

- With small sample sizes low and high variance can occur by chance
- Variance depends on expression level
- Try to remove (or minimize) the dependence of test statistics on variances (because small variance tend to lead to bigger test statistics).
- Solution: add a small constant to the denominator in calculating t statistics:

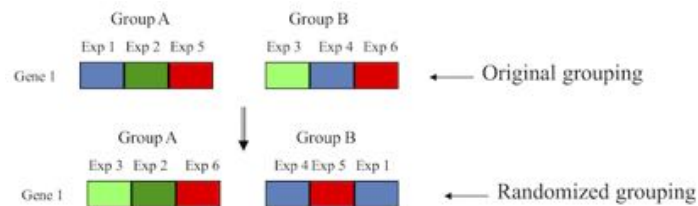
$$d_i = \frac{\bar{y}_i - \bar{x}_i}{s_i + s_0}$$

- \bar{y}_i, \bar{x}_i - Means of two groups for gene i.
- s_i - Standard deviation for gene i, assuming equal variance in both groups.
- s_0 - "Exchangeability factor" estimated using all genes.

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SAM two-class unpaired

- For each gene, compute the d-value (similar to a t-statistic). This is the observed d-value (d_i) for that gene.
- Randomly shuffle the expression values between groups A and B. Compute the d-value for each randomized set.
- Take the average of the randomized d-values for each gene. This is the 'expected relative difference' (d_E) of that gene. Difference between (d_i) and (d_E) is used to measure significance.
- Plot $d_{(i)}$ vs. $d_{E(i)}$
- Calculate FDR = average number of genes that exceed Δ in the permuted data.



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SAM statistics

- Define a statistic, based on the ratio of change in gene expression to standard deviation in the data for this gene.

$$d(i) = \frac{\bar{x}_I(i) - \bar{x}_U(i)}{s(i) + s_0}$$

← Difference between the means of the two conditions



$$s(i) = \sqrt{\left(\frac{\frac{1}{n_1} + \frac{1}{n_2}}{n_1 + n_2 - 2} \right) \left\{ \sum_m [x_m(i) - \bar{x}_I(i)]^2 + \sum_m [x_m(i) - \bar{x}_U(i)]^2 \right\}}$$

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Why s_0 ("fudge" factor)?

- Prevents $d_{(i)}$ from becoming too large when the variance is close to zero (which can lead to false positives)
- Choose one s_0 for the entire dataset, to make the coefficient of variation of $d_{(i)}$ approximately constant
- Typically, s_0 can be computed as the 90th percentile of the standard errors of all genes

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Estimating significance

- We have calculated a new statistics and we don't have a parametric description of the null distribution
- Solution: generate an empirical null distribution from a set of experiments where all hypotheses should be null
- Generate permutations of data labels so no difference is expected
- For each permutation p , calculate $d_{p(i)}$.

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Identifying Significant Genes

- Define a threshold Δ
- Find the smallest positive $d_{(i)}$ such that

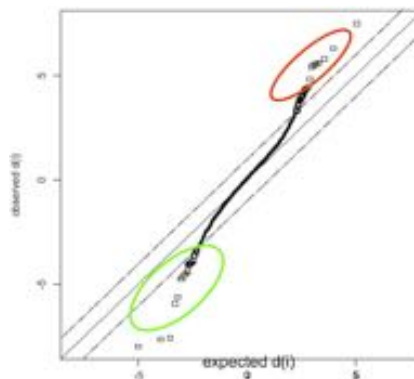
$$|d_{(i)} - d_{E(i)}| \geq \Delta$$

- Call it t_1
- In a similar manner, find the largest negative $d_{(i)}$. Call it t_2
- For each gene i , if $d_{(i)} \geq t_1 \vee d_{(i)} \leq t_2$, call it potentially significant

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Identifying Significant Genes

- Rank the original $d(i)$'s: $d_{(1)} \geq d_{(2)} \geq d_{(3)} \geq \dots$
- Plot $d_{(i)}$ vs. $d_{E(i)}$
- For most of the genes, $d_{(i)} \sim d_{E(i)}$



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Estimate FDR

- t_1 and t_2 will be used as cutoffs
- Calculate the average number of genes that exceed these values in the permutations.
- Estimate the number of falsely significant genes, under H_0 :

$$\frac{1}{n.perm} \sum_{p=1}^{n.perm} number\{d_{p(i)} \geq t_1 \vee d_{p(i)} \leq t_2\}$$

- Divide by the number of genes called significant

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Estimate FDR example

	$d(i)$	$d_p(i)$			
	8.3	8.3	8.4	7.9	8.1
t_1	4.2	3.2	4.4	2.5	1.6
	2.9	1.9	2.7	1.7	0.1
t_2	-0.5	0.3	-0.6	1.0	-2.1

$$FDR \approx \frac{7}{4} = 0.5833$$

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Estimate FDR from the reference distribution d

	$d(i)$	$d_p(i)$			
	8.3	8.3	8.4	7.9	8.1
t_1	4.2	3.2	4.4	2.5	1.6
	2.9	1.9	2.7	1.7	0.1
t_2	-0.5	0.3	-0.6	1.0	-2.1

$$FDR \approx \frac{7}{4} = 0.5833$$

Delta Δ is the half-width of the bar around the 45-degree line

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Other applications of SAM

- More than two groups
- Paired data
- Survival data, with censored response

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SAM summary

- Highly cited (>7000 citations), <http://www-stat.stanford.edu/~tibs/SAM/>.
- Implemented as Bioconductor package `siggenes`, and Excel plugin.
- Follow-up work: SAMSeq on RNA-seq DE test.
- Limitations: solutions for s_0 often sensitive to data.

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Summary on two-sample DE test

- Try to alleviate the “small sample variance” problem.
- Combine information from all genes.
- Many other variations of the model.
- In practice SAM and limma performs similarly.

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